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Rational Design of Ni(OH)₂ Hollow Porous Architecture for High-Sensitivity Enzyme-Free Glucose Sensor

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Abstract

Ni(OH)₂ electrocatalysts have acquired lots of research attentions as ideal substitutes for noble metals. However, their electrocatalytic performance still cannot meet the demands for applications due to the difficulties in electron transfer and mass transport. According to kinetics principle, the construction of hollow structure is regarded as an effective method to achieve outstanding electrocatalytic performance. In this work, Ni(OH)₂ hollow porous architecture (Ni(OH)₂ HPA) was simply synthesized through a coordinating etching and precipitating (CEP) method for the building of enzymatic-free glucose sensors. Ni(OH)₂ HPA presents large specific surface area (SSA), ordered diffusion channels, and structure stability. As a detection electrode for glucose, Ni(OH)₂ HPA exhibits eminent electroactivity in terms of high sensitivity (1843 $\mu\text{A mM}^{-1} \text{cm}^{-2}$), lower detection limit (0.23 μM), and short response time (1.4 s). The results demonstrate that Ni(OH)₂ HPA has practical applications for construction of enzymatic-free electrochemical sensors. The design of hollow structure also provides an effective engineering method for high-performance sensors.

Keywords: Ni(OH)₂, Hollow structure, Coordinating etching and precipitating, Glucose detection, Electrochemical sensor

Background

Glucose detection is very important in clinical biochemistry, food processing, and environmental monitoring. Developing a fast and reliable sensing method for glucose is an urgent demand for these applications [1–3]. Many techniques have been developed for this purpose, such as surface plasmon resonance [4], Fehling reagent method [5], optical rotation method [6], fluorescence [7], and electrochemistry [8]. Among these techniques, electrochemical methods have attracted more attentions owing to its high sensitivity, simplicity, low cost, and extraordinary low detection limit [9].

It is well known that the electrocatalytic activity of the working electrode determines the performance of the electrochemical sensors. Therefore, the design of electrode materials is vital for electrochemical sensors. Recently, transition metal hydroxides have been widely researched in this field due to the advantages of large reserves, low cost, and high activity derived from the

redox of metal composition [10]. Typically, Ni(OH)₂ was recognized as an ideal catalyst for glucose due to the redox couple (Ni³⁺/Ni²⁺) in alkaline medium. Although the metal components of Ni(OH)₂ can be used to restore the highly active electrons provided by the redox, their catalytic activity is still not high enough to meet the large-scale industrial production and people's living needs because of the difficulties in electron transfer and mass transport.

Inspired by the intimate connection between kinetics and microstructures (shape, size, component), scientists have already built different structured nanomaterials that are good for electrocatalytic dynamics, since the properties of the nanomaterials are usually structure-dependent [11]. Hollow porous nanostructure, which possesses well-defined interior voids, high specific surface area (SSA), low density, and structure stability, has attracted growing interests in recent years [12]. The available inner cavities effectively prevent active particles from aggregation and accommodate the structural strain accompanied with long-time measurements [13]. Otherwise, the functional shells can offer larger contact area between electrolyte and electrode, provide sufficient active sites, and

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reduce the length for both mass and electron transport [14]. Furthermore, the porous thin shell also supplies enough diffusion paths for analyte and intermediates, which are good for mass transport process [15]. In conclusion, high-active Ni(OH)₂ electrocatalysts can be acquired through the building of hollow porous feature.

Herein, cubic Ni(OH)₂ HPA is constructed by a Cu₂O-templated method inspired by the concept of coordinating etching and precipitating (CEP) route [16]. In order to demonstrate the advantages of hollow porous architecture, we comparatively evaluated the electrocatalytic activity of Ni(OH)₂ HPA and broken Ni(OH)₂ HPA (Ni(OH)₂ BHPA) through the detection of glucose. The hollow porous architecture provides larger SSA, more ordered transfer paths, and higher electron transfer efficiency compared to Ni(OH)₂ BHPA. Thus, the as-prepared Ni(OH)₂ HPA electrode presents higher electrocatalytic activity in terms of higher sensitivity, lower detection limit, and faster response time. The results demonstrate that Ni(OH)₂ HPA has potential applications for construction of electrochemical glucose sensors. This facile strategy also provides a valid method in the development of highly efficient nanomaterials for electrochemical sensors.

Methods/Experimental

Chemicals and Reagents

Copper chloride (CuCl₂·2H₂O; ≥ 99.0%), nickel chloride (NiCl₂·6H₂O; ≥ 98.0%), sodium thiosulfate (Na₂S₂O₃·5H₂O; ≥ 99.0%), polyvinyl pyrrolidone (PVP; *M_w* = 40,000), and sodium hydroxide (NaOH; ≥ 98.0%) were obtained from Chengdu Kelong. Glucose (Glu.; ≥ 99.5%), lactose (Lact.; ≥ 98.0%), sucrose (Sucr.; ≥ 99.5%), fructose (Fruc.; ≥ 99.0%), L-ascorbic acid (AA; ≥ 99.7%), uric acid (UA; ≥ 99.0%), and Nafion solution (5 wt% in mixture of lower aliphatic alcohols and water) were obtained from Sigma-Aldrich.

Synthesis of Ni(OH)₂ HPA

Firstly, cubic Cu₂O crystals were prepared following our previous work (Additional file 1: Figure S1) [17]. Then, 10 mg as-prepared cubic Cu₂O crystals and NiCl₂·6H₂O powder (4 mg) were dispersed into a mixed ethanol-water solution (10 mL, volume ratio = 1:1) by ultrasound. Thereafter, 0.33 g PVP powder was added with vigorous stirring for 0.5 h. Then, Na₂S₂O₃ (4 mL, 1 M) was added dropwise into the above system. The reaction was proceeded at normal temperature (25 °C) for 3 h. Finally, the products were washed for several times by centrifugation and dried at normal temperature. Ni(OH)₂ BHPA was obtained as contrast sample through strong ultrasonic treatment of Ni(OH)₂ HPA for 2 h (Additional file 1: Figure S2).

Materials Characterization

The crystal structure and composition of the products were measured by X-ray diffraction (XRD; Rigaku D/Max-2400) and X-ray photoelectron spectrometer (XPS; ESCALAB250Xi). The morphologies of the products were characterized by field emission scanning electron microscope (FESEM; FEI Quanta 250 and Zeiss Gemini 500) and high-resolution transmission electron microscope (HRTEM; FEI F20). The SSA and pore structure were analyzed on Brunauer-Emmett-Teller (BET; Belsort-max).

Electrochemical Measurements

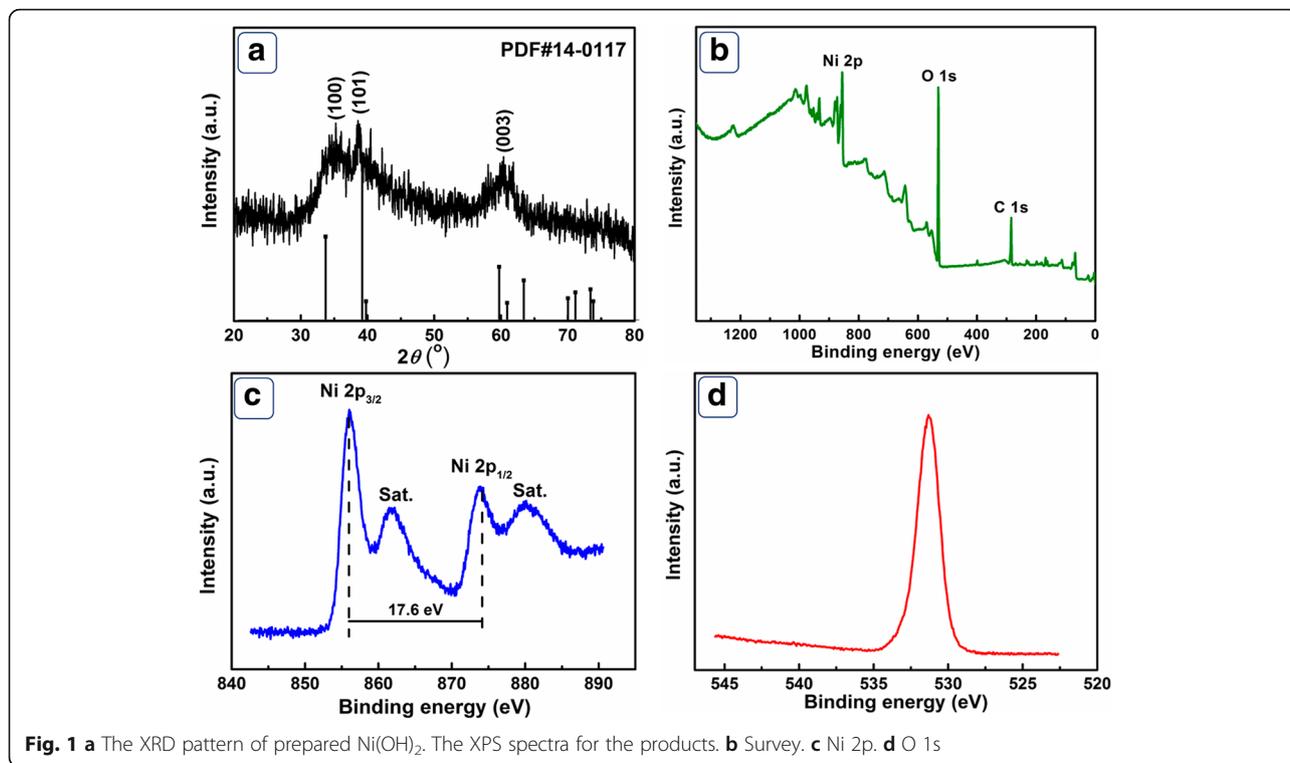
All electrochemical measurements were operated on electrochemical workstation (μIII Autolab). The working electrode is prepared by casting Nafion-impregnated Ni(OH)₂ HPA (or Ni(OH)₂ BHPA) powders onto a glassy carbon electrode (GCE; 3 mm in diameter) at room temperature. Specifically, 5 μL of the suspension (1 mg/ml in 0.05% Nafion solution) is dropped onto the pretreated GCE and dried by flowing N₂. A Pt foil and Ag/AgCl electrode were used as counter electrode and reference electrode, respectively. The electrocatalytic activity of working electrodes was measured by cyclic voltammetry (CV), chronoamperometry (CA), and electrochemical impedance spectroscopy (EIS). EIS data were collected between 0.01 and 100 kHz with a perturbation amplitude of 5 mV.

Results and Discussions

Characterizations

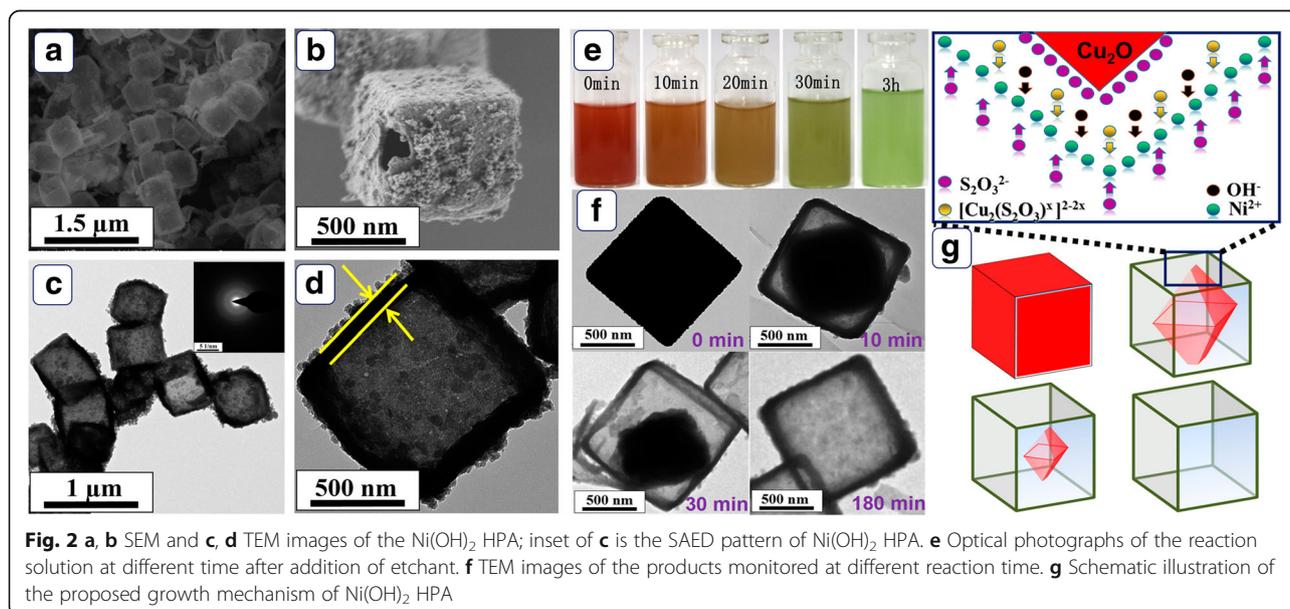
The XRD pattern of Ni(OH)₂ products was shown in Fig. 1a. The three main diffraction peaks can be assigned to (100), (101), and (003) crystalline planes of hexagonal β-Ni(OH)₂ (JCPDS no. 14-0117) [18]. The weak intensity of the diffraction peaks can be attributed to low crystallinity of the products. The purity and composition of the as-prepared Ni(OH)₂ were further investigated by XPS. The survey spectrum (Fig. 1b) demonstrates O 1s and Ni 2p peaks, revealing the main composition of the products. As displayed in Fig. 1c, the peaks located at 856.1 eV and 873.7 eV can be assigned to Ni 2p_{3/2} and Ni 2p_{1/2}, respectively. A binding energy separation of 17.6 eV is clearly observed, which is the characteristic of β-Ni(OH)₂. As displayed in Fig. 1d, the single peak at 531.2 eV corresponds to Ni–O–Ni bond in Ni–OH. By comparing the data with previous XPS studies, the presented Ni and O can be assigned to Ni²⁺ and OH[−] in Ni(OH)₂, respectively [16]. The analysis of XPS and XRD confirm the successful preparation of Ni(OH)₂ phase.

The low-magnification SEM image in Fig. 2a demonstrates a uniform cubic feature of the prepared Ni(OH)₂ products. The partly broken cube shown in Fig. 2b confirms the hollow characteristic of Ni(OH)₂ HPA.



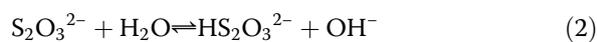
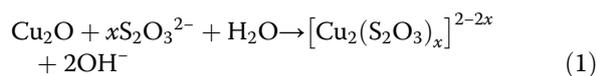
Moreover, the shell of Ni(OH)₂ HPA is built through the aggregation of numerous fine nanoparticles, making the shell rough and porous. The TEM images displayed in Fig. 2c further confirms the hollow structure of Ni(OH)₂ products. Meanwhile, no significant diffraction ring is observed in selected area electron diffraction (SAED)

pattern, suggesting low crystallinity of Ni(OH)₂ HPA. This result agrees well with the observation of XRD. Clearly investigated in Fig. 2d, the Ni(OH)₂ hollow cube has an edge length of ~ 600 nm and a shell thickness of ~ 50 nm. The hollow porous structure provides large SSA and amounts of diffusion channels, which may



benefit the mass diffusion process, resulting in satisfactory electrocatalytic activity.

The products prepared at different reaction stages were centrifuged and observed to realize the relevant formation principle. As observed in Fig. 2e, the color of the reaction system gradually turns light green and precipitates generate at the same time. As shown in Fig. 2f, the interior Cu_2O cores are gradually etched to octahedron after the addition of $\text{S}_2\text{O}_3^{2-}$ ions. The Cu_2O octahedrons finally disappear with the increase of reaction time. Combined with TEM images, the formation principle is illustrated in Fig. 2g. Apparently, $\text{S}_2\text{O}_3^{2-}$ ions adsorbed around Cu_2O cubes play versatile roles during the formation process of $\text{Ni}(\text{OH})_2$ HPA: (i) soluble $[\text{Cu}_2(\text{S}_2\text{O}_3^{2-})_x]^{2-2x}$ complex is formed through the combination of Cu^+ ions and $\text{S}_2\text{O}_3^{2-}$ (reaction (1)) and simultaneously OH^- ions are released. (ii) The hydrolysis of $\text{S}_2\text{O}_3^{2-}$ also releases OH^- ions (reaction (2)). (iii) Reactions (1) and (2) facilitate the formation of $\text{Ni}(\text{OH})_2$ (reaction (3)) [19]. Regarding the kinetics factors, the diffused OH^- ions from the interior determine the formation of $\text{Ni}(\text{OH})_2$ shell. Furthermore, the etching of Cu_2O is correlated to the transport of $\text{S}_2\text{O}_3^{2-}$ from exterior into internal space [20]. Synchronously controlling of OH^- and $\text{S}_2\text{O}_3^{2-}$ transport leads to the formation of well-defined $\text{Ni}(\text{OH})_2$ HPA.



The adsorption-desorption isotherm curve and the pore size distribution of $\text{Ni}(\text{OH})_2$ HPA and $\text{Ni}(\text{OH})_2$ BHPA are shown in Fig. 3. The SSA of $\text{Ni}(\text{OH})_2$ HPA is calculated to be $54.72 \text{ m}^2 \text{ g}^{-1}$ based on the desorption curve, which is much larger than that of $\text{Ni}(\text{OH})_2$ BHPA ($10.34 \text{ m}^2/\text{g}$). The decrease of SSA can be attributed to the destruction of

hollow structure and aggregation of the destroyed particles after ultrasonic treatment. The pore size distribution of $\text{Ni}(\text{OH})_2$ HPA and $\text{Ni}(\text{OH})_2$ BHPA both show regions below 10 nm, revealing the presence of nanopores between $\text{Ni}(\text{OH})_2$ nanoparticles. The pore size distribution of $\text{Ni}(\text{OH})_2$ HPA (inset of Fig. 3a) displays two concentrated regions of 20–40 nm and 60–85 nm, demonstrating the presence of micropores and mesopores. The micropores and mesopores might make the ion diffusion to active sites easier [21]. In the case of $\text{Ni}(\text{OH})_2$ BHPA (inset of Fig. 3b), a weak concentrated distribution is only investigated between 20 and 40 nm, indicating that the pore distribution of $\text{Ni}(\text{OH})_2$ BHPA is partly disordered. The decrease of SSA and destruction of ordered pore size may lead to difficulties in kinetics, resulting in poor electrocatalytic activity.

Electrochemical Measurements

The electrocatalytic activity of $\text{Ni}(\text{OH})_2$ HPA and $\text{Ni}(\text{OH})_2$ BHPA was studied through the detection of glucose in 0.1 M NaOH. Figure 4a shows the CVs of $\text{Ni}(\text{OH})_2$ HPA and $\text{Ni}(\text{OH})_2$ BHPA electrodes with and without 0.5 mM glucose. Obviously, the redox peak current of $\text{Ni}(\text{OH})_2$ HPA (curve I) is higher than $\text{Ni}(\text{OH})_2$ BHPA (curve III) due to larger SSA. Upon addition of 0.5 mM glucose, the current responses of $\text{Ni}(\text{OH})_2$ HPA electrode (curve II) are higher than $\text{Ni}(\text{OH})_2$ BHPA electrode (curve IV). Otherwise, $\text{Ni}(\text{OH})_2$ HPA electrode shows lower onset potential (0.41 V) than that of $\text{Ni}(\text{OH})_2$ BHPA electrode (0.44 V). The higher electrocatalytic activity of $\text{Ni}(\text{OH})_2$ HPA can be attributed to high electron transfer rate, large SSA, and ordered pore structure provided by the hollow porous architecture. The electrocatalysis of glucose on $\text{Ni}(\text{OH})_2$ HPA electrode is driven by $\text{Ni}(\text{OH})_2/\text{NiOOH}$ redox couple in alkaline medium on the basis of following reactions [22], and the corresponding schematic diagram is illustrated in Scheme 1.

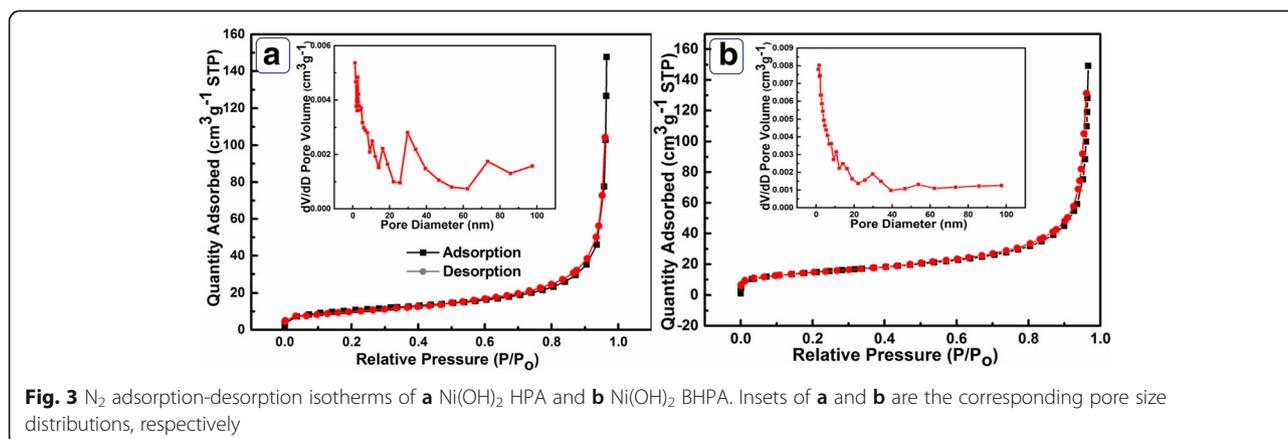
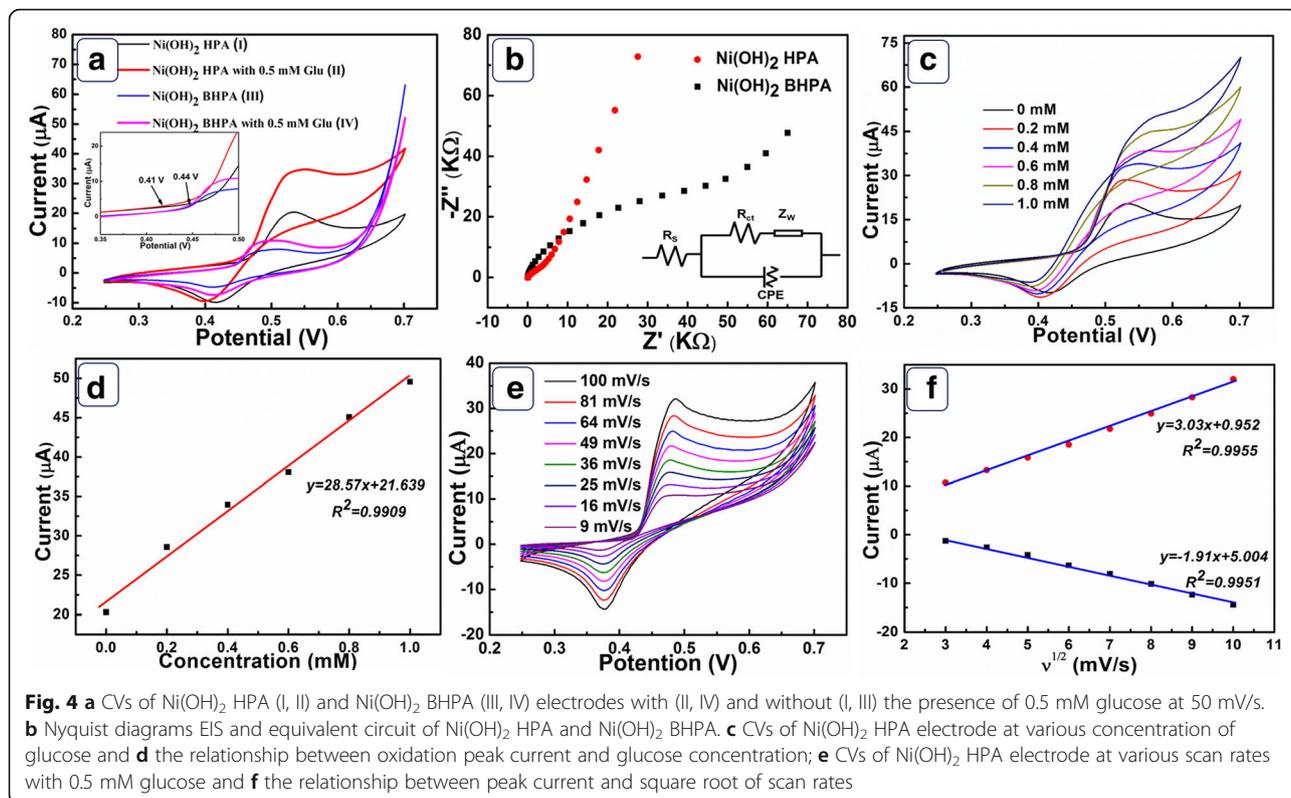
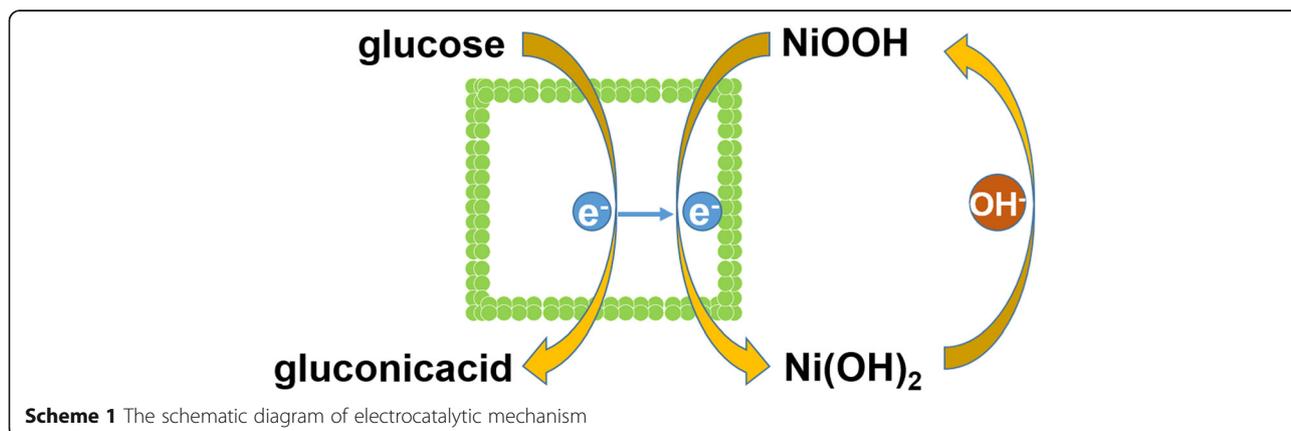


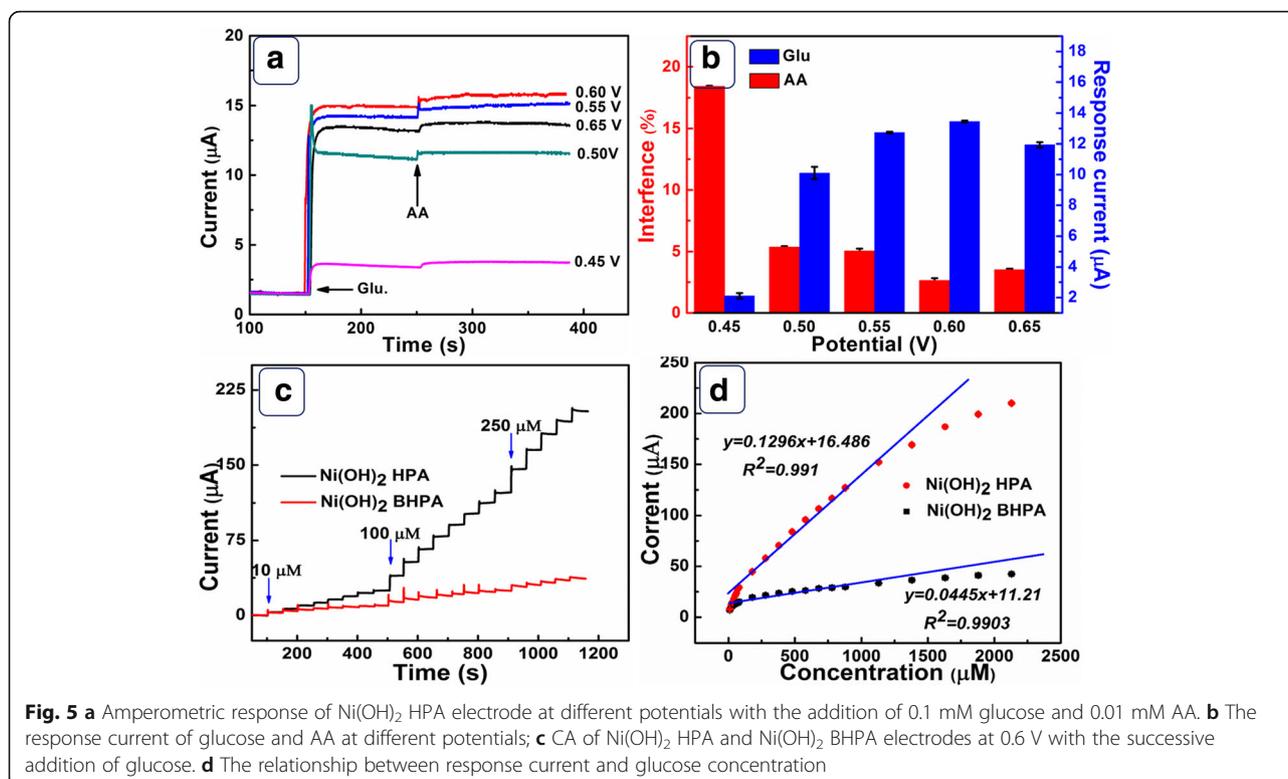
Fig. 3 N_2 adsorption-desorption isotherms of **a** $\text{Ni}(\text{OH})_2$ HPA and **b** $\text{Ni}(\text{OH})_2$ BHPA. Insets of **a** and **b** are the corresponding pore size distributions, respectively



In order to confirm the kinetic advantages of hollow porous feature, EIS spectra of Ni(OH)₂ HPA and Ni(OH)₂ BHPA were measured (Fig. 4b). The EIS spectrum is characterized by a semicircle in high-frequency and an incline in low-frequency region. As shown in Additional file 1: Table S1, Ni(OH)₂ HPA electrode exhibits smaller internal

resistance (*R_s*) and electron transfer resistance (*R_{ct}*) than Ni(OH)₂ BHPA. Moreover, the Warburg impedance (*Z_w*) of Ni(OH)₂ HPA is larger than that of Ni(OH)₂ BHPA, indicating more effective mass transfer rate. The difficulties in mass transfer kinetics of Ni(OH)₂ BHPA can be ascribed to the destruction of ordered diffusion channels and aggregation of the broken cubes. In conclusion, Ni(OH)₂ HPA electrode exhibits advantages in both electron and mass transfer kinetics compared to Ni(OH)₂ BHPA. Figure 4c is the CVs of Ni(OH)₂ HPA electrode in 0.1 M NaOH with different glucose concentration at





50 mV/s. The oxidation peak current linearly increases with the glucose concentration (Fig. 4d), revealing applications in electrochemical glucose sensors. The CVs of Ni(OH)₂ HPA electrode with 0.5 mM glucose under different scan rates were recorded in Fig. 4e. As shown in Fig. 4f, the peak current linearly depends on the square root of scan rates, revealing diffusion controlled electrochemical process.

In order to confirm the optimized working potential, the current response of glucose and the interference of AA were considered at different potentials (Fig. 5a). From the statistics data displayed in Fig. 5b, Ni(OH)₂ HPA electrode exhibits minimum interference to AA and maximum current response to glucose at 0.6 V. Thus, 0.6 V was selected as the optimized working potential. Figure 5c shows the amperometric response curves of Ni(OH)₂ HPA and Ni(OH)₂ BHPA electrodes at 0.6 V. Ni(OH)₂ HPA electrode presents more sensitive response to glucose than Ni(OH)₂ BHPA electrode. Figure 5d is the corresponding calibration curves of Ni(OH)₂ HPA and Ni(OH)₂ BHPA electrodes. For Ni(OH)₂ HPA electrode, the results show a good linear region between 0.08 mM and 1.13 mM. The fitting equation is $y = 0.1296x + 16.486$ ($R^2 = 0.991$). By accurate calculation, Ni(OH)₂ HPA electrode has a sensitivity of $1843 \mu\text{A mM}^{-1} \text{cm}^{-2}$, which is higher than Ni(OH)₂ BHPA electrode ($632 \mu\text{A mM}^{-1} \text{cm}^{-2}$). The detection

limit of Ni(OH)₂ HPA electrode is calculated to be $0.23 \mu\text{M}$ ($S/N = 3$), which is lower than Ni(OH)₂ BHPA ($0.67 \mu\text{M}$). As displayed in Additional file 1: Figure S3, Ni(OH)₂ HPA electrode presents a shorter response time (1.4 s) compared to Ni(OH)₂ BHPA electrode (1.8 s). The analytical performances of Ni(OH)₂ HPA electrode are compared with other Ni(OH)₂-based electrodes, and the data are listed in Table 1. Notably, Ni(OH)₂ HPA electrode presents higher electroactivity towards glucose in terms of high sensitivity, low detection limit, and rapid response, indicating great potential applications as an electrochemical glucose detection electrode.

Common interferences in human blood, including Lact., Suct., Fruct., UA, and AA, are involved to evaluate the selectivity of Ni(OH)₂ HPA electrode [23]. As displayed in Fig. 6a, no more than 3.8% interferences are observed for all the interferences. The second current response for glucose retains 98.1% of its first signal. Figure 6b displays the amperometric response of Ni(OH)₂ HPA electrode towards 0.1 mM glucose within 2400 s at 0.6 V. The final response signal still retains approximately 93.5% of its original data, revealing an excellent long-term stability of Ni(OH)₂ HPA electrode. In Fig. 6c, current responses for one Ni(OH)₂ HPA electrode were tested for ten times. The signals display a relatively standard deviation (RSD) of 4.8%, demonstrating outstanding reproducibility. Moreover, the five Ni(OH)₂ HPA electrodes exhibit a satisfying RSD of 5.3%

Table 1 Comparison of researched electrode with reported nonenzymatic glucose sensors based on Ni(OH)₂

Electrode	Sensitivity ($\mu\text{A mM}^{-1} \text{cm}^{-2}$)	Detection limit (μM)	Response time (s)	Reference
Ni(OH) ₂ HPA/GCE	1843	0.23	1.4	This work
Ni(OH) ₂ -CNT ^a -PVDF composite	–	23	3	[22]
Ni(OH) ₂ /nanoflowers	265.3	0.5	3	[24]
Ni(OH) ₂ /TiO ₂	192.5	8	1	[25]
Ni(OH) ₂ NP/MoS _x	162	5.8	2	[26]
Nano Ni(OH) ₂ platelet-like	202	6	5–7	[27]
Ni(OH) ₂ /ECF ^b	1342.2	0.1	3	[28]
Ni(OH) ₂ -graphene	494	0.6	2	[29]
Ni(OH) ₂ @oPPyNW ^c	1049.2	0.3	10	[30]
Ni(OH) ₂ /Au/GCE	371.2	0.92	4	[31]

^aCarbon nanotubes^bElectrospun carbon nanofiber^cOver-oxidized polypyrrole nanowires

(Fig. 6d). Ni(OH)₂ HPA electrode possesses excellent selectivity, satisfying stability and reproducibility, demonstrating attractive applications in electrochemical glucose sensors.

Conclusions

We have used a facile strategy inspired by the CEP principle to controllably fabricate uniform Ni(OH)₂ HPA at room temperature. Ni(OH)₂ HPA presents large SSA, ordered diffusion channels, and high structure stability. As a

electrochemical detection electrode for glucose, Ni(OH)₂ HPA exhibits higher sensitivity of $1843 \mu\text{A mM}^{-1} \text{cm}^{-2}$, faster response time (1.4 s), and lower detection limit of $0.23 \mu\text{M}$ compared to broken sample (1.8 s, $0.67 \mu\text{M}$). The Ni(OH)₂ HPA electrode presents improved electrochemical sensing performance towards glucose, revealing promising feature for the practical analytical application. The hollow porous architecture is also confirmed as an effective strategy to obtain high-performance electrocatalysts.

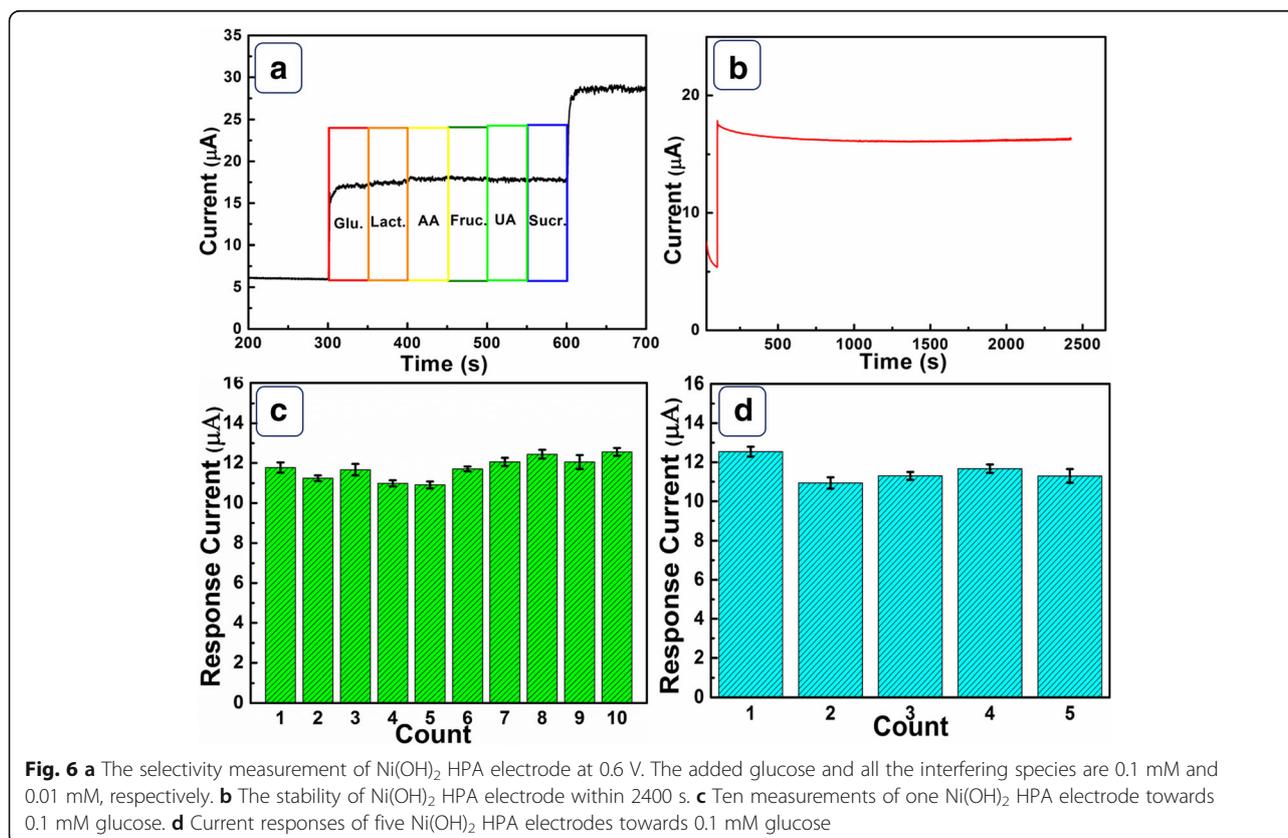


Fig. 6 **a** The selectivity measurement of Ni(OH)₂ HPA electrode at 0.6 V. The added glucose and all the interfering species are 0.1 mM and 0.01 mM, respectively. **b** The stability of Ni(OH)₂ HPA electrode within 2400 s. **c** Ten measurements of one Ni(OH)₂ HPA electrode towards 0.1 mM glucose. **d** Current responses of five Ni(OH)₂ HPA electrodes towards 0.1 mM glucose

Additional file

Additional file 1: Figure S1. (a) SEM and (b) XRD pattern of the prepared Cu_2O . Figure S2 The SEM image of $\text{Ni}(\text{OH})_2$ BHPA. Figure S3 The detection limit of $\text{Ni}(\text{OH})_2$ HPA and $\text{Ni}(\text{OH})_2$ BHPA. Table S1 Comparison of researched $\text{Ni}(\text{OH})_2$ HPA electrode with $\text{Ni}(\text{OH})_2$ BHPA about EIS. (DOCX 1575 kb)

Abbreviations

AA: L-ascorbic acid; BET: Brunauer-Emmett-Teller; CA: Chronoamperometry; CEP: Coordinating etching and precipitating; CNT: Carbon nanotubes; ECF: Electrospun carbon nanofiber; EIS: Electrochemical impedance spectroscopy; FESEM: Field emission scanning electron microscope; Fruc.: Fructose; GCE: Glassy carbon electrode; Glu.: Glucose; HPA: Hollow porous architecture; Lact.: Lactose; $\text{Ni}(\text{OH})_2$ BHPA: Broken $\text{Ni}(\text{OH})_2$ HPA; oPPyNW: Over-oxidized polypyrrole nanowires; PVP: Polyvinyl pyrrolidone; *R*_{ct}: Electron transfer resistance; *R*_s: Internal resistance; RSD: Relatively standard deviation; SAED: Selected area electron diffraction; SSA: Specific surface area; Sucr.: Sucrose; UA: Uric acid; XPS: X-ray photoelectron spectrometer; XRD: X-ray diffraction; *Z*_w: Warburg impedance

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Availability of Data and Materials

The datasets are available without restriction.

Authors' Contributions

LLT design the experiment and wrote the paper. GGH and YCY did the electrochemical measurements. XB prepared the materials and made the characterizations. MJC and JBW provided financial supports. All authors read and approved the final manuscript.

Competing Interests

The authors declare that they have no competing interests.

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